Treatment of Scars: A Review

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Hypertrophic Scars and Keloids

The complex interplay of events that occurs following injury to the skin does not always eventuate in a normal, smooth skin surface. Rather, the skin often responds to injury with a proliferation of fibrous tissue. When tissue response to injury is overzealous, the result is a hypertrophic scar or keloid.

By definition, hypertrophic scars remain within the boundaries of the original wound, whereas keloids extend beyond the original area of skin injury (Figs 1 and 2). Some authors include a temporal component to differentiate these two types of lesions as well, suggesting that a hypertrophic scar that is present for greater than 12 months and with margins that extend beyond the original site of the wound evolves into a keloid [1]. The time between cutaneous injury and the onset of hypertrophic scar or keloid formation may vary from weeks to years, but hypertrophic scars differ from keloids in their tendency to regress spontaneously over time, while keloids remain elevated [2]. Despite these definitions, hypertrophic scars and keloids are difficult to differentiate in many circumstances due to the fact that their clinical characteristics often overlap.

Histologically, keloids and hypertrophic scars are composed of thick, hyalinized collagen bundles arranged in nodules consisting of small fibrocytes and separated by bands of immature fibroblasts [3]. There are multiple microvessels that often appear occluded by an excess of endothelial cells [4]. Keloids have additionally been shown to contain an increased amount of hyaluronidase.

Many different theories have been proposed to explain the array of clinical presentations of scars. Complex biological systems, including biochemical, metabolic, and immunological, have been implicated in hypertrophic scar and keloid formation [5–8]. However, because of the interdependence of the steps involved in the wound-healing cascade, it has been impossible to separate the events sufficiently to implicate any one cell type or blood-borne factor as being responsible for keloid and hypertrophic scar formation.

Wound healing is arbitrarily divided into three phases: inflammation, granulation tissue formation, and matrix remodeling. The first phase, initiated by neutrophils, involves a well-organized cascade of inflammatory cells. Later, macrophages, which may be the most important cells in wound healing, elaborate a variety of cytokines that influence the wound environment and eventuate in granulation tissue formation. Finally, fibroblasts migrate into the area, proliferate, and recapitulate ontogeny by depositing new collagen—first as type III and later as type I. Simultaneously, angiogenic factors released into the wound result in the formation of new capillaries. Under ordinary circumstances, scar maturation occurs due to the regression of stimulatory factors. Coincident with scar maturation is a decrease in angiogenic stimuli and, therefore, a decrease in the hyperemia associated with early wound repair. Collagen synthesis and remodel-

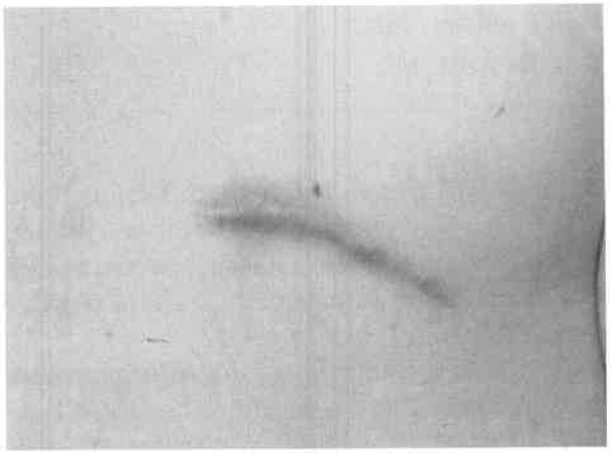


Fig 1. Hypertrophic scar within the borders of the original incision.

Fig 2. Keloid of the ear, showing extension of the scar beyond the initial ear-piercing wound.

ing also normalize within 6 to 12 months of the initial tissue injury. While scars never regain 100% of their original tensile strength, they eventually achieve approximately 70% to 80% of the strength of normal skin.

Various factors, including ethnic background and location of injury, are thought to be important in determining whether a scar heals in a cosmetically acceptable manner or develops into a keloid or hypertrophic scar. Hypertrophic scars and keloids affect approximately 4.5% to 16% of the black and hispanic population. Caucasians are less susceptible, with a white-to-black susceptibility ratio estimated at 1:3.5 to 1:15 [9]. Keloids result from an inherited metabolic alteration in collagen [10]. Both types of scars occur most commonly on the back, chest, shoulders, and earlobes. Other high-incidence areas include the proximal upper limbs, the pectoral areas, and the lower face [11]. They may arise in areas that have been traumatized (e.g., burns, surgery, acne, vaccinations), but may also occur spontaneously, especially on the anterior aspect of the chest. They are most commonly observed between 10 and 30 years of age [2, 9, 12]. Clinically, hypertrophic scars occur less frequently and are usually milder in severity after sutured surgical incisions than they are after burns or other wounds in which epithelialization is delayed. Scars can also be more severe following poorly designed surgical wound closures (with excess tension placed on the wound) and following wound infection. Hypertrophic scars usually occur within the first 6 to 8 weeks after skin reepithelialization and undergo a rapid growth phase that can last up to 6 months, with gradual "maturation" over the next 1 to 2 years [13]. In some patients they may gradually disappear, while in others they continue to enlarge and may last a lifetime [14]. The excess collagen produced in these abnormal wounds is probably related to oversynthesis rather than to an increase in the absolute number of fibroblasts. The resulting overproduction of collagen surpasses collagen degradation during the remodeling phase of wound healing, resulting in hypertrophy [15]. Investigators have demonstrated decreased expression of collagenase in hypertrophic scar fibroblasts, suggesting a possible etiological mechanism for the excessive accumulation of collagen in hypertrophic scars [16].

The presence of a keloid or hypertrophic scar is frequently cosmetically unacceptable to the affected individual. In addition, it may be painful or pruritic and may restrict range of motion. Physicians have attempted to improve the appearance of scars by both physical and chemical means for many years. Various methods of manipulation have been enlisted to normalize their topography as well as to eliminate erythema and dyspigmentation. Because the treatment of scars is often undertaken for, at least in part, cosmetic concerns, it must be free of adverse sequelae in addition to being effective.

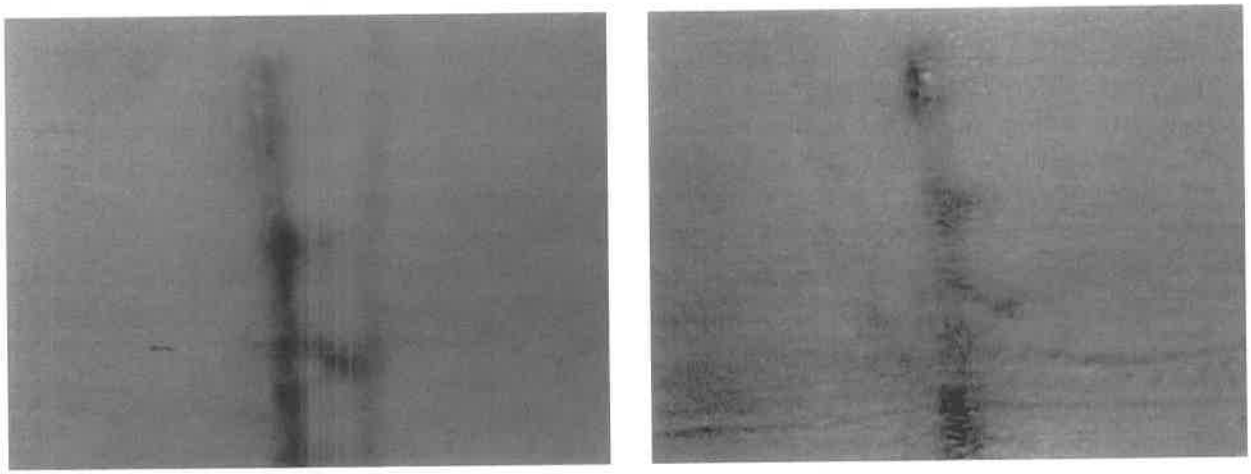
There is no universally accepted treatment modality resulting in permanent hypertrophic or

keloid scar ablation. Surgical excision is usually followed by recurrence unless concomitant prophylactic measures are employed, because the new surgical wound is prone to the same mechanical, immunological, and biochemical forces as the original scar. Recurrence rates following excisional surgery alone have been estimated at 45% to 100% [17].

Proposed management or prevention of scars include three distinctly different therapeutic approaches: (1) correction of abnormal collagen metabolism when the equilibrium between collagen synthesis and degradation has been destroyed, (2) alteration of the immune/inflammatory response, and (3) manipulation of the mechanical properties of wound repair [18]. Systemic and local pharmacological treatments have been advocated for keloids and hypertrophic scars that take into account these three approaches. Colchicine has been shown to affect collagen metabolism at three points: inhibition of collagen synthesis by disruption of microtubular systems, stimulation of collagenase, and interference with wound contraction by a direct effect on the myofibroblasts [15, 19]. Antihistamines have been instituted in the treatment of hypertrophic scars based on the observation that mast cells may play a role in scar formation [20]. Lathyrogens, such as D penicillamine and beta aminopropionitrile, prevent collagen cross-linking after its secretion from the fibroblast, increasing its vulnerability to enzymatic degradation. Local methods of manipulation have included topical and intralesional steroids [21, 22], topical application of retinoic acid [23, 24], intralesional hyaluronidase [25], topical zinc [26], topical vitamin E [27], topical putrescine to modify type III collagen cross-linking [28], surgical revision [29], radiation therapy [30, 31], pressure therapy [32], silicone gel sheeting [10, 33–37], cryotherapy [38–40], and laser treatment [41–60]. Most of these treatments are associated with high recurrence rates and several may lead to significant side effects or be painful or inconvenient, resulting in decreased patient compliance.

Intralesional injection of corticosteroid has been a cornerstone of both treatment and prophylaxis of hypertrophic scars and keloids [21, 61–63]. When surgery is combined with intradermal corticosteroids the recurrence rate in the majority of studies falls below 50% [17]. The effect of corticosteroids may be explained in part by interruption of the inflammatory response [1]. The specific mechanism of action of corticosteroids is related to both suppression of collagen synthesis and to the release of collagenase inhibition resulting in collagen catabolism. Once considered the standard treatment for hypertrophic sternal scars [63], intralesional injection with triamcinolone solution has shown clinical efficacy but has also been associated with multiple side effects including atrophy, white beadlike skin deposits, pain on injection, and pigmentary changes in up to 63% of patients [64]. In addition, intralesional corticosteroid injection may result in the development of telangiectasias overlying the treated area.

The molecular basis for the excessive fibrosis that results in the formation of keloids and hypertrophic scars has not been delineated. However, investigators have begun to study the role of products of the inflammatory response and the regulation of collagen synthesis in scar formation. Interleukin-1, produced by monocytes, increases collagen and fibronectin synthesis by fibroblasts. In contrast, recombinant human interferon gamma has been shown to downregulate collagen synthesis and may be useful in the treatment of diseases characterized by collagen overproduction [65]. Based on data from Kahan and colleagues, who used recombinant human interferon gamma to treat 10 patients with systemic sclerosis [66], Larrabee and associates used this lymphokine to treat 10 patients with hypertrophic scars and keloids. Larrabee et al reported moderate clinical and symptomatic improvement in patients with existing abnormal scars, however the most dramatic result was achieved in a patient who was successfully prophylaxed with interferon gamma following keloid excision. Side effects, including headache, reversible granulocytopenia, and elevation of hepatic transaminase levels, are dose- and route-dependent [1]. The probable mechanism of action of interferon gamma in downregulating collagen is through reduction in cellular messenger ribonucleic acid (mRNA) [67, 68]. In contrast to steroids, which produce a general reduction in the inflammatory response, recombinant human interferon gamma targets a specific site of action in the fibroblast



A B

Fig 3. (A, B) Hypertrophic scar on the abdomen (A) before and (B) after 6 months of nightly silicone gel sheeting.

that reduces collagen production. Whether there is an excess of interleukin-1 or a deficiency in interferon gamma in individuals with a tendency to develop abnormal scars has not been determined.

Growth factors known to modulate wound healing include basic fibroblastic growth factor, the BB isoform of platelet-derived growth factor, insulinlike growth factor-I (IGF-I) and transforming growth factor beta (TGF-β) [69]. Recently it has been suggested that IGF-I-induced suppression of collagenase mRNA and activity may be a mechanism by which IGF-I promotes the development of postburn hypertrophic scars [69]. TGF-β is released by platelets at the site of injury and is highly chemotactic for macrophages and monocytes. It also stimulates production of collagen and fibronectin by fibroblasts, probably by enhancing the effect of epidermal growth factor on the fibroblastic population, stimulating epithelial differentiation and initiating deoxyribonucleic acid (DNA) and RNA synthesis [71–73]. Roberts and colleagues have suggested that excess production or abnormal sensitivity to this ubiquitous growth factor may be important in pathologic fibrosis, not only in wounds but also in other systemic fibrotic disorders [74].

Since the first report by Perkins and associates [73] in 1982, various investigators have demonstrated significant scar improvement following application of topical silicone gel sheeting for 2 to 4 months [10, 13, 34, 74–78] (Fig 3). Topical application of silicone gel to scars for at least 12 hours daily has been shown not only to provide early relief of symptoms and to enhance patient compliance due to the lack of pain associated with the treatment [64], but may also prevent hypertrophic and keloid scar formation [79]. Katz [78] demonstrated that 79% of scars treated immediately after reepithelialization (following full-thickness surgical revision and scar abrasion) with silicone sheeting did not recur within 6 months.

The effect of silicone gel has been shown to be unrelated to pressure [75], but may depend on scar hydration instead [35–37]. Recent studies using occlusive silicone creams have confirmed that silicone gel sheets function through scar hydration and occlusion [35–37, 80]. Phillips and colleagues [80] demonstrated a significant reduction in pruritus and pain, and an increased pliability within hypertrophic scars, but not keloids, treated with hydrocolloid dressing or moisturizer for 2 months. Scar pigmentation and elevation, however, remained unchanged [81]. While the mechanism by which hydration exerts its effects on scars is unknown, the reduction in water vapor loss is postulated to decrease capillary activity, thereby reducing collagen deposition and scar hypertrophy [82]. Wood and coworkers [83] have reported that occlusion decreases interleukin-1α mRNA. Interleukin-1 is a proinflammatory cytokine exhibiting pleiotropic effects that include a direct increase in fibroblastic collagenase glycosaminoglycan synthesis. This cytokine also increases the production of interleukin-6,

which in turn activates human fibroblastic synthesis of extracellular matrix components [84]. It has yet to be determined whether silicone-based or nonsilicone-based sheeting is more effective in the treatment of scars.

A continuous pressure of approximately 80 mmHg has been shown to elongate and flatten some scars due to remodeling of scar collagen [84]. The mechanism through which continuous pressure reduces the size and thickness of keloids and hypertrophic scars is unknown, but may be related to a demonstrated reduction of intralesional mast cell numbers and consequent reduction in histamine production [86]. Local tissue hypoxia has also been proposed as the mode of action through which continuous pressure exerts its effects, as compressed keloids have significantly reduced tissue oxygen (pO_2) levels and increased tissue carbon dioxide (pCO_2) levels when compared with normal tissue [87].

Physical manipulation of keloids and hypertrophic scars has been attempted with the use of cryotherapy. The therapeutic effects of cryotherapy are related to direct cell damage as well as to changes in the microcirculation initiated by freezing, which leads to stasis of blood and subsequent dermal anoxia. Finally, occlusion of vessel lumina with thrombi produces tissue necrosis and sloughing. The collagen fibers that subsequently form are flat and normal in quantity, with axes parallel to the skin surface [40]. Treatment usually involves two or three freeze-thaw cycles at each of the 2 to 10 treatment sessions. Layton and Colleagues [38] reported that cryosurgery was more effective than intralesional triamcinolone in the treatment of acne keloids. Keloids on the face, as well as those that were grossly palpable, however, showed a poor response to both treatment modalities. Those keloids that demonstrated greater blood flow by Doppler, and that were also more clinically erythematous and of more recent onset, responded better to both treatments compared with less vascular lesions. This study was flawed by its short follow-up of only 8 weeks as well as its limitation of each treatment to two sessions. Zouboulis and coworkers [39] in their randomized prospective study of 93 white patients with keloids and hypertrophic scars, found improved responses in those patients treated with three or more sessions compared with individuals treated once or twice. They reported excellent responses in 32.3%, good responses in 29%, and no response in 9.7%. Responders exhibited no progressions or recurrences in an average follow-up period of 32 months [39]. These results may be biased in terms of their applicability to keloids and hypertrophic scars in general because the authors excluded individuals with darker skin tones who are known to demonstrate a higher recurrence rate. Several authors have reported that cryotherapy offers no better results than simple surgical excision [3, 88]. Rusciani and associates [40] suggest that the reported lack of efficacy may be due to inadequate freeze times or the failure to continue treatment when the results of the initial freeze are less than outstanding. In their series, complete flattening was achieved in 73% of 65 scars and there were no recurrences at an average follow-up of 31 months. Side effects were limited to hypopigmentation and light to moderate atrophy in 3 patients. The hypopigmentation due to the cold sensitivity of melanocytes is largely permanent and renders cryotherapy less desirable in dark-skinned patients. The postoperative phase may be disturbing to the patient because cryotherapy produces an open wound that takes several weeks to heal [40]. Better results may be achieved with active keloids of less than a 2-year duration because collagen synthesis and degradation is typically normalized to levels similar to those of normal scars beyond that time [89]. In addition, pedunculated lesions are thought to respond better than those on a broad base due to the more limited vascular supply of the former [40].

Excision followed by radiation therapy has been touted as a useful and effective method of keloid eradication [31, 90–94]. The effectiveness of radiation therapy is thought to relate to its inhibition of neovascular buds and proliferating young fibroblasts associated with early wound healing, resulting in a decreased amount of collagen production [95]. Klumpar and coworkers [96] found that the electron beam offers no advantage over orthovoltage radiation after comparing the two modalities in the treatment of 186 patients over 17 years. Doornbos and associates [90] analyzed a large series of patients treated over a 25-year period and concluded that the total radiation dose is more significant than the timing of

treatment initiation, the size of the fraction given, the duration of treatment, or the location of the keloid. The authors recommended a dose of at least 1,500 cGy [90]. Radiation therapy has traditionally been associated with the development of side effects such as local hypo- and hyperpigmentation, erythema, telangiectasia, and atrophy. Recently, Sclafani and colleagues [30] reported only a 12.5% incidence of earlobe keloid recurrence when radiation was used as an adjunct to excision, while adjunctive steroid injection resulted in a recurrence rate of 33%. While these authors noted no alteration of skin pigmentation, wound dehiscence, chronic dermatitis, or neoplastic change, controversy continues over whether to treat benign lesions with radiation therapy due to the perceived risk of adverse long-term sequelae. It has been suggested that radiation is not practical for use in children or in areas of potential carcinogenesis including the breast and thyroid gland [1, 12]. However, only two cases of malignancy have been reported in patients who underwent excision followed by radiation therapy and both tumors were thought to be unrelated to the treatment [97]. Radiotherapy may also be used in the treatment of established keloids, although recurrence rates are higher than when radiation is delivered after excision [31, 90].

Bailin [98] first reported good results in a series of patients with keloids excised by the continuous-wave CO_2 laser in 1982. The laser's beneficial effect was attributed to its nontraumatic and anti-inflammatory properties [98]. In 1991 Norris [58] further evaluated the efficacy of CO_2 laser excision as a primary modality for the treatment of keloids. Of 31 patients retrospectively studied, only 1 subject was free of recurrence while 9 other patients required intralesional steroids to suppress recurrence and 13 others were judged to be treatment failures [58]. It was concluded that CO_2 laser excision of keloids fails to suppress their growth and recurrence. This conclusion was later corroborated by Apfelberg and associates [54] in 1989 when 7 patients with nine keloids, located on the trunk, nuchal region, back, and earlobe, were treated by CO_2 laser excision. Only 1 patient, after earlobe keloid excision, experienced marked improvement, which was maintained for 9 months, but required the continuous use of pressure earrings [54]. Following their experience treating a patient with a large truncal keloid, Olbricht and Arndt [99] concluded that CO_2 laser excision of keloids should be reserved for special situations such as large or draining keloids that require debulking before the institution of other measures to control regrowth.

Since 1983 when Anderson and Parrish [100] published their landmark paper describing the theory of selective photothermolysis, laser technology has exploded with the development and refinement of a myriad of high-energy, target-specific pulsed and scanned laser systems. Prior to that time, lasers were less selective and often resulted in nonspecific thermal damage to tissue surrounding the treated area. The newer high-energy pulsed and scanned lasers have revolutionized the treatment of many skin disorders by the precise targeting of specific tissue components resulting in a vast reduction of scarring as a sequela of treatment.

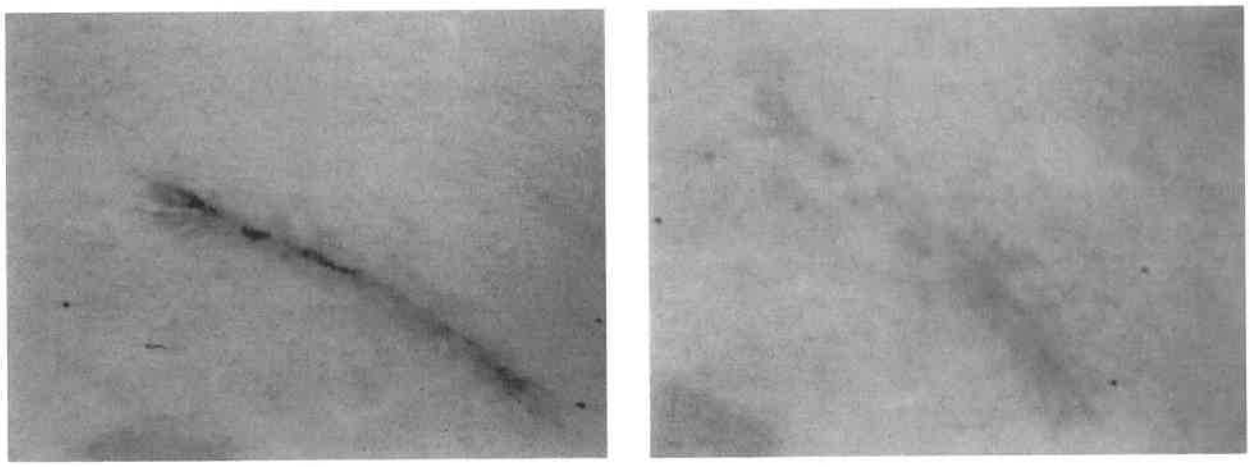
The argon laser was one of the first lasers applied to the treatment of keloids and hypertrophic scars. Ginsbach and Kohnel [101] in 1978 were the first to report encouraging results with the argon laser in the treatment of hypertrophic scars. The argon laser was thought to work by coagulation of the capillary plexus leading to an area of localized anoxia with the production of lactic acid by glycolysis. The resulting decreased pH was believed to lead to granulocytic lysis with the release of enzymes, including collagenase. This increase in tissue levels of collagenase was postulated to be a direct result of laser treatment, but was also thought to be an indirect result of a decrease in $alpha_2$ macroglobulin, a collagenase inhibitor. The resulting increase in collagenase, by either mechanism, would result in increased collagenolysis, which would then lead to the flattening of hypertrophic scars [55, 101]. However, another study in 1984 failed to corroborate the efficacy of the argon laser, with only approximately one half of 82 patients achieving a good to excellent objective response following argon (77 patients) and CO_2 (5 patients) laser treatment. No symptomatic improvement associated with the scars was seen in 80% of treated patients within 14 days postoperatively [55]. Apfelberg and associates [53] divided keloids into thirds for the purpose of comparing argon and CO_2 laser irradiation to control (nontreated) sites in 13

patients with keloids of 5.6 years average duration. While several of the treated patients experienced temporary symptomatic improvement, only one maintained a response at the 6-month follow-up [53]. Hulsbergen-Henning and colleagues [57] showed similar results with only 3 of 45 patients treated with the argon laser demonstrating scar improvement (decreased color and size) of more than 50%. These authors postulated that the mechanism of action of the argon laser was related to heat conduction causing shrinkage of dermal tissue, but that the treatment was superficial, resulting in only temporary scar improvement [57].

The neodymium:yttrium-aluminum-garnet (Nd:YAG) laser (1064 nm, continuous wave) has been shown to exert an effect on collagen metabolism. Initial studies showed decreased collagen content in Nd:YAG laser-treated pig skin [101]. Later, collagen production was shown to be selectively inhibited by a direct photobiological effect of this laser, while DNA replication and cell viability of fibroblasts were unaffected [56]. A follow-up study documented 2 patients, each followed for 3 years, in whom treatment of keloids with the Nd:YAG laser resulted in significant flattening, softening, reduction in size, and normalization of color. The authors postulated that perhaps even more effective keloid eradication could be achieved by a combination of conventional excision or CO_2 laser vaporization of excess tissue followed by Nd:YAG laser treatment to suppress collagen production [56]. The Nd:YAG laser appeared to work by infarction with subsequent charring and sloughing of the irradiated area, which was then left to heal by secondary intention. A deterrent to the use of lasers in the visible or near-infrared spectrum is the presence of large amounts of melanin in the epidermis because laser energy is absorbed by the pigment at a superficial level, resulting in reduced thermal injury to the deeper target tissue. The Nd:YAG, argon, and CO_2 lasers can each temporarily decrease collagen synthesis, thus providing a distinct advantage over scalpel excision, which cannot. Keloids remain a distinct problem, however, as they represent a permanent change in physiology manifested as an inability to regulate a balance between collagen synthesis and lysis in wound healing [59].

Persistent erythema in scars is the result of excess vasculature. Increased vascularization results in fibroblast proliferation and excess collagen accumulation. Over the past 10 years great strides have been made with the use of the 585-nm vascular-specific flashlamp-pumped pulsed dye laser in the treatment of hypertrophic and/or erythematous scars and keloids. However, the exact mechanism whereby hypertrophic and keloid scars are altered by the 585-nm laser system remains unknown. The first study demonstrating an improvement in scars using the 585-nm pulsed dye laser was reported by Alster and colleagues [47] in 1993. Ten argon laser-induced scars within port wine stains received five laser treatments over a 10-month period. Using optical profilometry to measure skin surface texture, the authors demonstrated a return of normal skin markings after 585-nm irradiation, representing a change in the surface texture of the treated scars to more closely resemble that of normal skin. The improvement was manifested either as flattening of the hypertrophic portions of the scars and/or the reappearance of skin surface markings in the scarred areas. This study represented the first use of an objective parameter (optical profilometry) to assess the degree of improvement in scar texture as an adjunct to subjective evaluation by observation and photography. Histological examination demonstrated that the dilated vascular channels present in scar tissue resolved following laser treatment in 8 of 10 patients but remained unaltered in 2 others. The improvement in the laser-treated scars persisted at the 6-month study follow-up examination and Alster [43] later reported that no recurrence or worsening of the scars occurred in the 4 years following treatment.

In a subsequent study of 14 patients with erythematous scars, Alster [41] demonstrated a 57% to 83% improvement with the 585-nm flashlamp pulsed dye laser following one or two treatments respectively (Fig 4). The clinical improvement, which included softening and flattening of the scars as well as decreased erythema, persisted at the 6-month follow-up examination [41]. More recently, Alster and McMeekin [49] showed that hypertrophic facial acne scars demonstrated significant clinical and textural improvement within one to two laser sessions using the 585-nm



A B

Fig 4. (A, B) Hypertrophic scar on the breast (A) before and (B) 6 weeks after a second treatment with a 585-nm pulsed dye laser.

pulsed dye laser. The average energy densities used in these studies ranged from 6.0 to 7.0 J per square centimeter with a 5-mm spot size.

The first controlled study of the response of hypertrophic scars and keloids to the 585-nm flashlamp-pumped pulsed dye laser was performed on median sternotomy scar halves by Alster and Williams [51] in 1995. The authors objectively demonstrated the difference between laser-treated and untreated scar tissue by analyzing skin surface textures, histological specimens, erythema and height measurements, pliability scores, and symptomatology. The significant improvement observed in each of these study parameters within the laser-irradiated scar halves persisted for at least 6 months following treatment.

In an uncontrolled study, Dierickx and associates [52] treated 15 patients with erythematous/hypertrophic scars refractory to various traditional treatment modalities with the 585-nm pulsed dye laser. The authors reported that an average of 1.8 laser treatments at fluences ranging from 6.0 to 7.5 J per square centimeter delivered through a 5-mm spot size produced an average improvement of 77%. Scars less than 1 year old tended to respond more readily to treatment than older scars, as did facial scars in comparison to those on the extremities or buttocks. While the initial objective of the study was to improve the color of the scars, the authors reported subjective softening and flattening as an unexpected consequence of laser therapy [52]. Similar conclusions were reached by Goldman and Fitzpatrick [103], who treated 48 patients presenting with erythematous/hypertrophic scars with the 585-nm pulsed dye laser with or without concomitant intralesional steroids. Patients who received combination treatment with simultaneous administration of intralesional steroids (5 to 10 mg per milliliter) and laser treatment appeared to achieve greater resolution than those treated with the laser alone, although it was unclear from the report exactly which patients received the combination treatment [103].

Initially, the 585-nm pulsed dye laser was directed at the vascular component of scars to reduce or eliminate persistent erythema, with the softening and flattening of the treated scars as an incidental finding. There is no consensus on the mechanism of improvement in scar texture produced by this vascular-specific laser. Some plausible theories include a decrease in cellular nutrition and function within the irradiated scar through the laser's vascular specificity, leading to decreased microvascular perfusion, an alteration of the ratio of collagen metabolism resulting in catabolism as the overriding force by laser-induced tissue hypoxia, superheating of collagen fibers resulting in dissociation of disulfide bonds, and subsequent organized realignment of collagen fibers [52]. Because fibroblastic growth and other factors are known to be affected by histamine, another likely explanation for the effective-

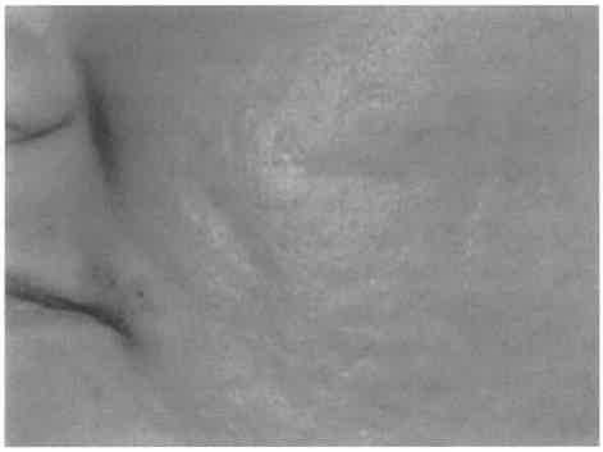


Fig 5. Atrophic acne scars on the face.

ness of the 585-nm laser may be related to the increase in regional mast cell numbers observed after laser irradiation [51].

In terms of patient selection for the treatment of scars with the 585-nm pulsed dye laser, individuals with pale skin phototypes (I or II) are the best candidates because the increased amount of melanin in darker skin competes with hemoglobin for absorption of the laser energy. The most appropriate timing of treatment has not yet been determined. While scars may improve spontaneously during the first 6 to 12 months after integumental injury, especially in terms of erythema, anecdotal evidence suggests that earlier treatment of scars, within the first month following surgery or trauma, may prevent hypertrophy in individuals who are keloid prone [43, 46, 104, 105]. Perhaps the initiation of laser treatment within the first few weeks following injury may prevent or arrest scar proliferation so that fewer laser sessions may be required to achieve the desired clinical outcome [42, 44, 106].

Atrophic Scars

Atrophic scarring can be a result of surgery, trauma, and such common conditions as acne vulgaris and varicella (Fig 5). Many different procedures, alone and in combination, have been employed to correct these dermal depressions, which are disfiguring and may even be exacerbated by makeup due to its enhancement of textural variation.

Dermabrasion, a manual technique for removing the epidermis and a portion of the dermis that utilizes an abrasive wheel driven by a high-speed rotary engine, has long been considered the definitive treatment for atrophic and pitted scars secondary to acne [104]. The physiological basis of healing after dermabrasion rests on the capacity of the skin to regenerate its epidermal cover by epithelialization through skin appendages, primarily the pilosebaceous units. Reepithelialization takes 7 to 10 days and erythema may last for an additional 2 to 12 weeks. There is evidence that improvement in the clinical appearance of abraded skin correlates with histological evidence of new collagen formation in the dermis [107, 108]. The wider and more shallow the pits, the better the cosmetic results obtained with dermabrasion [109]. For correction of icepick or other well-demarcated scars, the results obtained with dermabrasion are enhanced when used in combination with punch excision and primary closure and/or punch grafting. During the excision or grafting, icepick scars are isolated and removed either by primary closure or replacement with a postauricular graft. The scars are then dermabraded 4 to 8 weeks later [110, 111]. Dermabrasion has also been performed in conjunction with chemical peeling for enhanced clinical results. The technique is highly operator dependent and is limited by various factors including poor visualization during surgery due to bleeding and the potential for transmission of infectious particles. Complications of dermabrasion include permanent hypopigmentation with a noticeable line of demarcation between treated and untreated skin, transient hyperpigmentation, infection, and milia formation [112].

In addition to excision [109] or punch grafting [113, 114], other surgical procedures have been implemented in the correction of atrophic acne scarring. Subcision is a method of cutting under a depressed scar or contour using a tribeveled hypodermic needle, which is inserted under the skin through a needle puncture. Once beneath the defect, the surgeon attempts to raise its base manually to the level of the surrounding skin surface. The effectiveness of this technique, which may have to be repeated several times for each scar, is thought to be related to both surgical release of the fibrous strands anchoring the scar

and spontaneous fibroplasia induced by injury [115].

Tissue augmentation for depressed scars has been performed with a variety of filling materials including collagen [116–118], silicone [119], fat [120], fibrin [121, 122], and Fibrel [123]. Silicone, while proving successful in the recontouring of soft-tissue defects, produced an unacceptably large percentage of complications. Bovine collagen injection provides a relatively risk-free correction of dermal defects following adequate skin-testing procedures; however, treatment must be repeated at frequent intervals. Currently, investigators are studying various methods of harvesting and cultivating autologous collagen to avoid allergic reactions to the injected material [124]. In addition, this autologous material should remain in place for an extended duration because it is not as readily targeted by enzymes or antibodies. Fibrotic or icepick scars are not amenable to treatment with filler substances. Pinski and Roenigk [120] reported their results following autologous fat transplantation in 43 patients followed over 3 to 48 months. Cosmetic defects treated included linear morphea, expression lines, acne scars, discoid lupus erythematosus scars, and posttraumatic scars. The authors noted that the nature of the cosmetic defect was the most important factor in determining graft longevity. The greatest amount of fat resorption was encountered in treating fibrotic acne scars. Only approximately 30% of injected fat was present following 12 months. The authors proposed that the impaired vascularity of these lesions contributed to the decreased viability of the fat grafts, and postulated that the fibrosis process and not true fat cell survival was responsible for the contour improvement seen after fat transplantation. Complications included temporary swelling and minor bruising at the recipient site, and minor tenderness at the donor site [120]. There has also been a report of unilateral blindness following transplantation of autologous fat to the glabella [125].

The recent development of high-energy, pulsed, or scanned CO_2 lasers utilizing high peak powers and short pulse durations diminishes thermal conduction to normal skin structures by limiting the pulse duration to a period of time shorter than the thermal relaxation time of water-containing tissue [126–128]. The advantage of the newest, high-energy pulsed or scanned CO_2 lasers in comparison with more conventional forms of treatment lies in their ability to vaporize precisely predictable and reproducible amounts of tissue with each progressive laser pass. In a histological study comparing depth of ablation of laser resurfacing, dermabrasion, and chemical peels, Fitzpatrick and colleagues [129] demonstrated that dermal vaporization plus necrosis depth secondary to CO_2 laser resurfacing was directly proportional to pulse energy as well as the number of laser passes. Therefore, with the use of the high-energy, pulsed CO_2 laser the operator is able to remove as little or as much tissue as desired depending on the clinical situation, a distinct advantage over other resurfacing techniques [129]. Similar to dermabrasion, the mechanism of CO_2 laser resurfacing involves the regeneration of epidermis and dermis from residual appendageal structures as well as collagen remodeling, which may continue for at least 1 year following treatment. Treatment with the CO_2 laser also exerts a thermal effect on collagen that results in collagen shrinkage and may be responsible for clinical improvement following even a relatively superficial ablation [130]. Lastly, an increase in myofibroblasts within laser-induced wounds has been discovered that may provide a contracted scaffold for subsequent collagen organization [131].

Garrett and associates [132] were among the first to report cosmetically acceptable results in the treatment of acne scarring with the CO_2 laser utilizing the continuous and superpulsed modes. The computer assisted scanner that was available at that time, however, produced a depression in its footprint [132]. Later, in a report of 13 patients with skin types III and IV who underwent CO_2 laser resurfacing for acne scars, 10 patients were treated with the UltraPulse laser and 3 were treated with the Sharplan Silktouch flashscanner [133]. The authors treated the "shoulders" of scars first with one to four laser passes, followed by one laser pass to the whole cosmetic unit or vice versa. A 25% average improvement following one laser treatment was reported, with saucer-shaped scars responding better than icepick scars. These authors postulated that deeper scars would possibly show greater response to punch

A B

Fig 6. (A, B) Atrophic acne scars (A) before and (B) 6 months after high-energy, pulsed CO_2 laser resurfacing treatment.

excision or punch grafting prior to laser resurfacing. Apfelberg [134] reached similar conclusions after treating 13 patients with mild and severe acne scarring with the UltraPulse CO_2 laser over a 24-month period [134]. All patients were treated with three to five laser passes using energies of 300 to 500 mJ. He concluded that laser resurfacing of severe atrophic acne scars achieves only moderate results, while laser treatment of mild acne can provide excellent results.

In another study that included 50 patients with skin phototypes I through V and moderate to severe atrophic facial acne scars treated with a high-energy, pulsed CO_2 laser, blinded assessor ratings of clinical improvement averaged 81.4% (Fig 6). Two to five laser passes over the scarred areas were delivered using a 3-mm collimated handpiece. Skin texture analyses of the laser-irradiated scars demonstrated a return of normal skin surface markings and were comparable with those analyses obtained in normal adjacent skin. All patients were followed for 6 months with no evidence of scar recurrence or worsening [135]. On the contrary, a continued clinical improvement in atrophic scars has been observed for at least 1 year following the laser resurfacing procedure [136]. This phenomenon is probably related to ongoing collagen remodeling, which begins during the final phase of wound healing and during which the dermis responds to injury with the production of collagen and matrix proteins [137, 138]. Similar clinical results are achieved following laser resurfacing of traumatic scars.

Fortunately, side effects are uncommon following high-energy pulsed or scanned CO_2 laser surgery. Erythema, typically lasting 4 weeks to 4 months and sometimes 6 to 9 months, is expected and should not be regarded as a complication of treatment. Prolonged erythema with tissue induration and tenderness usually is indicative of early hypertrophic scarring. As has been reported with dermabrasion, the risk of scarring may be increased in patients who have been treated with oral retinoids within the preceding 2 years as well as in those patients who develop postoperative bacterial or herpetic infection [139–141]. Milia and acne may occur temporarily, especially if occlusive ointments are used postoperatively in acne-prone skin. Hyperpigmentation is not uncommon, occurring in 30% or more of patients within 4 to 6 weeks postoperatively. Although patients with olive skin tones are more likely to hyperpigment, this side effect can occur in any patient undergoing cutaneous laser resurfacing [46, 135]. Although most cases of hyperpigmentation will eventually resolve spontaneously, the fading process typically takes several months. Hypopigmentation, a common sequela of dermabrasion, has been reported infrequently as a delayed complication of laser resurfacing. It is usually not seen until several months following the laser procedure and seems to occur more

commonly in areas treated with a greater number of laser passes. In addition, areas predisposed to hypopigmentation include the oral commissures and areas of skin that have been previously treated by other modalities including dermabrasion and phenol peels. Hypopigmentation that has been commonly associated with dermabrasion may be attributed to deeper follicular melanocytic injury than what routinely occurs during laser resurfacing alone [133]. While a series of superficial chemical peels may improve the overall appearance of the skin surrounding these areas of hypopigmentation, the pigment loss appears to be permanent.

Laser resurfacing represents a major advance in the treatment armamentarium for the improvement of atrophic facial scarring. The technique is of tremendous importance to patients who suffer from the sequelae of some earlier inflammatory condition or physical trauma. While continuous-wave CO_2 lasers have been utilized for tissue vaporization for many years, their use for cutaneous resurfacing was limited by heat conduction to surrounding tissues with resultant scarring. With the advent of high-energy pulsed and scanned CO_2 laser technology, precisely controlled, layer-by-layer tissue vaporization may be achieved with minimal thermal damage to adjacent normal skin when the correct laser parameters and techniques are employed. This advance in laser technology has simplified performance of cutaneous resurfacing while minimizing adverse sequelae to the patient.

Summary

Scars are notoriously difficult to treat. Multiple treatments have been advocated in the past with varying degrees of success. Hypertrophic scars and keloids have been shown to respond to pressure therapy, intralesional steroids, systemic chemotherapy, radiation, topical silicone, and laser treatment. Simple vaporization or mechanical destruction of these proliferative scars often leads to recurrence or scar worsening.

Atrophic scars, on the other hand, respond favorably to controlled vaporization or mechanical procedures including high-energy CO_2 laser irradiation, dermabrasion, and chemical peel. More predictable and controlled depths of dermal destruction can be achieved by proper use of the latest available lasers.

References

1 Larrabee WF, East CA, Jaffe HS, et al. Intralesional interferon gamma treatment for keloids and hypertrophic scars. Arch Otolaryngol Head Neck Surg 1990;116:1159–1161
2 Datubo-Brown DD. Keloids: a review of the literature. Br J Plast Surg 1990;43:70–77
3 Muti E, Ponzio E. Cryotherapy in the treatment of keloids. Ann Plast Surg 1983;11:227–232
4 Kischer CW, Thies AC, Chvapil M. Perivascular myofibroblasts and microvascular occlusion in hypertrophic scars and keloids. Hum Pathol 1983;13:819–824
5 Cracco C, Stella M, Teich Alasia S, Filogamo G. Comparative study of Langerhans cells in normal and pathological human scars. II. Hypertrophic scars. Eur J Histochem 1992;36:53–65
6 Rockwell WB, Cohen IK, Ehrlich HP. Keloids and hypertrophic scars: a comprehensive review. Plast Reconstr Surg 1989;84:827–837
7 Scott PG, Dodd CM, Tredget EE, et al. Chemical characterization and quantification of proteoglycans in human post-burn hypertrophic and mature scars. Clin Sci 1996; 90:417–425
8 Zhang LQ, Laato M, Muona P, et al. Normal and hypertrophic scars: quantification and localization of messenger RNAs for type I, III and VI collagens. Br J Dermatol 1994;130:453–459
9 Ketchum LD, Cohen IK, Masters FW. Hypertrophic scars and keloids. Plast Reconstr Surg 1974;53:140–154
10 Gold MH. Topical silicone gel sheeting in the treatment of hypertrophic scars and keloids: a dermatologic experience. J Dermatol Surg Oncol 1993;19:912–916
11 O'Sullivan ST, O'Shaughnessy M. Aetiology and management of hypertrophic scars and keloids. Ann R Coll Surg Engl 1996;78:168–175
12 Cosman B, Crikelair GF, Ju DM, et al. The surgical treatment of keloids. Plast Reconstr Surg 1961;27:335–358
13 Ahn ST, Monafo WW, Mustoe TA. Topical silicone gel: a new treatment for hypertrophic scars. Surgery 1989;4: 781–787
14 Fulton JE. Silicone gel sheeting for the prevention and management of evolving hypertrophic and keloid scars. Dermatol Surg 1995;21:947–951
15 Cohen IK, Diegelmann RF. The biology of keloid and hypertrophic scar and the influence of corticosteroids. Clin Plast Surg 1977;4:297–299
16 Arakawa M, Hatamochi A, Mori Y, et al. Reduced collagenase gene expression in fibroblasts from hypertrophic scar tissue. Br J Dermatol 1996;134:863–868
17 Berman B, Bieley HC. Adjunct therapies to surgical management of keloids. Dermatol Surg 1996;22:126–130
18 Cohen IK, McCoy BJ. The biology and control of surface overhealing. World J Surg 1980;4:289–295
19 Peacock EE. Pharmacologic control of surface scarring in human beings. Ann Surg 1981;193:592–597
20 Topol BM, Lewis VL, Benveniste K. The use of antihistamine to retard the growth of fibroblasts derived from

human skin, scar and keloid. Plast Reconstr Surg 1981; 68:227–232
21 Griffith VH. The treatment of keloids with triamcinolone acetonide. Plast Reconstr Surg 1966;38:202–208
22 Vallis CP. Intralesional injection of keloids and hypertrophic scars with the Dermo-jet. Plast Reconstr Surg 1967; 40:255–262
23 Nelson DL, Balian G. The effect of retinoic acid on collagen synthesis by human dermal fibroblasts. Coll Relat Res 1984;4:119–128
24 Janssen de Limpens AMP. The local treatment of hypertrophic scars and keloids with topical retinoic acid. Br J Dermatol 1980;103:319–323
25 Cornbleet T. Treatment of keloids with hyaluronidase. JAMA 1954;154:1161–1163
26 Soderberg T, Hallmans T, Bartholdson L. Treatment of keloids and hypertrophic scars with adhesive zinc tape. Scand J Plast Reconstr Surg 1982;16:261–266
27 Edgerton MT, Hanrahan EM, Davis WB. Use of vitamin E in the treatment of keloids. Plast Reconstr Surg 1951;8: 224–227
28 Dolynchuk KN, Ziesmann M, Serletti JM. Topical putrescine (Fibrostat) in treatment of hypertrophic scars: phase II study. Plast Reconstr Surg 1996;97:117–123
29 Engrav LH, Gottlieb JR, Millard SP, et al. A comparison of intramarginal and extramarginal excision of hypertrophic burn scars. Plast Reconstr Surg 1988;81:40–43
30 Sclafani AP, Gordon L, Chadha M, Romo T. Prevention of earlobe keloid recurrence with postoperative corticosteroid injections versus radiation therapy. Dermatol Surg 1996;22:569–574
31 Borok TL, Bray M, Sinclair I, et al. Role of ionizing irradiation for 393 keloids. Int J Radiat Oncol Biol Phys 1988;15:865–870
32 Linares HA, Kischer CW, Dobrovsky M, Larson DL. On the origin of the hypertrophic scar. J Trauma 1973;13: 70–75
33 Gold MH. A controlled clinical trial of topical silicone gel sheeting in the treatment of hypertrophic scars and keloids. J Am Acad Dermatol 1994;30:506–507
34 Mercer NSG. Silicone gel in the treatment of keloid scars. Br J Plast Surg 1989;42:83–87
35 Sawada Y, Sone K. Treatment of scars and keloids with a cream containing silicone oil. Br J Plast Surg 1990;43: 683–688
36 Sawada Y, Sone K. Beneficial effects of silicone cream on grafted skin. Br J Plast Surg 1992;45:105–108
37 Sawada Y, Sone K. Hydration and occlusion treatment for hypertrophic scars and keloids. Br J Plast Surg 1992; 45:599–603
38 Layton AM, Yip J, Cunliffe WJ. A comparison of intralesional triamcinolone and cryosurgery in the treatment of acne keloids. Br J Dermatol 1994;130:498–501
39 Zouboulis CC, Blume U, Buttner P, Orfanos CE. Outcome of cryosurgery in keloids and hypertrophic scars. Arch Dermatol 1993;129:1146–1151
40 Rusciani L, Rosse G, Bono R. Use of cryotherapy in the treatment of keloids. J Dermatol Surg Oncol 1993;19: 529–534
41 Alster TS. Improvement of erythematous and hypertrophic scars by the 585 nm pulsed dye laser. Ann Plast Surg 1994;32:186–190
42 Alster TS. Laser treatment of erythematous/hypertrophic and pigmented scars in 26 patients—a discussion. Plast Reconstr Surg 1995;95:91–92
43 Alster TS. Laser treatment of hypertrophic scars. Fac Plast Surg Clin North Am 1996;4:267–274
44 Alster TS. Laser treatment of scars. In: Alster TS, Apfelberg DB, eds. Cosmetic laser surgery. New York: John Wiley, 1996;81–92
45 Alster TS. Laser treatment of hypertrophic scars, keloids, and striae. Dermatol Clin 1997;15:419–429
46 Alster TS. Manual of cutaneous laser techniques. Philadelphia: Lippincott Raven, 1997
47 Alster TS, Kurban AK, Grove GL, et al. Alteration of argon laser-induced scars by the pulsed dye laser. Lasers Surg Med 1993;13:368–373
48 Alster TS, Lewis AB. Use of a high-energy, pulsed CO_2 laser singly and in combination with a 585 nm pulsed dye laser in the treatment of scars. Lasers Surg Med Suppl 1996;8:31–32 [abstract]
49 Alster TS, McMeekin TO. Improvement of facial acne scars by the 585 nm flashlamp-pumped pulsed dye laser. J Am Acad Dermatol 1996;35:79–81
50 Alster TS, Nanni CA. Pulsed dye laser treatment of hypertrophic burn scars. J Am Acad Dermatol 1997 [in press]
51 Alster TS, Williams CM. Treatment of keloid sternotomy scars with the 585 nm flashlamp-pumped pulsed dye laser. Lancet 1995;345:1198–1200
52 Dierickx C, Goldman MP, Fitzpatrick RE. Laser treatment of erythematous/hypertrophic and pigmented scars in 26 patients. Plast Reconstr Surg 1995;95:84–90
53 Apfelberg DB, Maser MR, Lash H, et al. Preliminary results of argon and carbon dioxide laser treatment of keloid scars. Lasers Surg Med 1984;4:283–290
54 Apfelberg DB, Maser MR, White DN, Lash H. Failure of carbon dioxide laser excision of keloids. Lasers Surg Med 1989;9:382–388
55 Henderson DL, Cromwell TA, Mes LG. Argon and carbon dioxide laser treatment of hypertrophic and keloid scars. Lasers Surg Med 1984;3:271–277
56 Abergel RP, Dwyer RM, Meeker CA, et al. Laser treatment of keloids: a clinical trial and an in vitro study with Nd:YAG laser. Lasers Surg Med 1984;4:291–295
57 Hulsbergen-Henning JP, Roskam Y, van Gemert MJ. Treatment of keloids and hypertrophic scars with an argon laser. Lasers Surg Med 1986;6:72–75
58 Norris JE. The effect of carbon dioxide laser surgery on the recurrence of keloids. Plast Reconstr Surg 1991;87: 44–49
59 Sherman R, Rosenfeld H. Experience with the Nd:YAG laser in the treatment of keloid scars. Ann Plast Surg 1988;21:231–235
60 Stern JC, Lucente FE. Carbon dioxide laser excision of earlobe keloids. A prospective study and critical analysis of existing data. Arch Otolaryngol Head Neck Surg 1989; 115:1107–1111
61 Ketchum LD, Robinson DW, Masters FW. Follow-up on treatment of hypertrophic scars and keloids with triamcinolone. Plast Reconstr Surg 1971;48:256–259
62 Murray RD. Kenalog and the treatment of hypertrophic scars and keloids in negroes and whites. Plast Reconstr Surg 1963;31:275–280
63 Maguire HC. Treatment of keloids with triamcinolone acetonide injected intralesionally. JAMA 1956;192:325–329
64 Sproat JE, Dalcin A, Weitauer N, Roberts RS. Hypertrophic sternal scars: silicone gel sheet versus kenalog injection treatment. Plast Reconstr Surg 1992;90:988–992
65 Pittet B, Rubbia-Brandt L, Desmouliere A, et al. Effect of gamma-interferon on the clinical and biologic evolution

of hypertrophic scars and Dupuytren's disease: an open pilot study. Plast Reconstr Surg 1994;93:1224–1235
66 Kahan A, Amor B, Menkes CJ, Strauch G. Recombinant interferon gamma in the treatment of systemic sclerosis. Am J Med 1989;87:272–277
67 Duncan MR, Berman B. Gamma interferon is the lymphokine and beta interferon the monokine responsible for inhibition of fibroblast collagen production and late but not early fibroblast proliferation. J Exp Med 1985;162: 516–527
68 Granstein RD, Murphy GF, Margolis RJ, et al. Gamma interferon inhibits collagen synthesis in vivo in the mouse. J Clin Invest 1987;79:1254–1258
69 Hathaway CL, Arnold AM, Rand RP, et al. Differential expression of IGFBP's by normal and hypertrophic scar fibroblasts. J Surg Res 1996;60:156–162
70 Ghahary A, Shen YJ, Nedelec B, et al. Collagenase production is lower in post-burn hypertrophic scar fibroblasts than in normal fibroblasts and is reduced by insulin-like growth factor-1. J Invest Dermatol 1996;106: 476–481
71 Russell SB, Trupin KP, Rodriguez-Eaton S, et al. Reduced growth-factor requirement of keloid-derived fibroblasts may account for tumor growth. Proc Natl Acad Sci USA 1988;85:587–591
72 Roberts AB, Sporn MB, Assoian RK, et al. Transforming growth factor type β: rapid induction of fibrosis and angiogenesis in vivo and stimulation of collagen formation in vitro. Proc Natl Acad Sci USA 1986;83:4167–4171
73 Wahl SM, Hunt DA, Wakefield LM, et al. Transforming growth factor type beta induces monocyte chemotaxis and growth factor production. Proc Natl Acad Sci USA 1987;84:5788–5792
74 Perkins K, Davey RB, Walis KA. Silicone gel: a new treatment for burns and contractures. Burns 1982;9:406–410
75 Quinn KJ, Evans JH, Courtney JM, et al. Non-pressure treatment of hypertrophic scars. Burns 1985;12:102–108
76 Quinn KJ. Silicone gel in scar treatment. Burns 1987;13: 533–540
77 Carney SA, Cason CG, Gowar JP, et al. Cica-Care gel sheeting in the management of hypertrophic scarring. Burns 1994;20:163–167
78 Dockery GL, Nilson RZ. Treatment of hypertrophic and keloid scars with silastic gel sheeting. J Foot Ankle Surg 1994;33:110–119
79 Katz BE. Silicone gel sheeting in scar therapy. Cutis 1995;56:65–67
80 Bieley HC, Berman B. Effects of a water-impermeable, non-silicone-based occlusive dressing on keloids. J Am Acad Dermatol 1996;35:113–114
81 Phillips TJ, Gerstein AD, Lordan V. A randomized controlled trial of hydrocolloid dressing in the treatment of hypertrophic scars and keloids. Dermatol Surg 1996;22: 775–778
82 Davey RB, Wallis KA, Bowering K. Adhesive contact media—an update on graft fixation and burn scar management. Burns 1991;17:313–319
83 Wood LC, Elias PM, Sequeira-Martin SM, et al. Occlusion lowers cytokine mRNA levels in essential fatty acid-deficient and normal mouse epidermis, but not after acute barrier disruption. J Invest Dermatol 1994;103: 834–838
84 Duncan MR, Berman B. Stimulation of connective tissue related biosynthetic functions of cultured human adult dermal fibroblasts by recombinant human interleukin 6. J Invest Dermatol 1991;97:686–692
85 Kischer CW, Shetlar MR, Shetlar CL. Alteration of hypertrophic scars induced by mechanical pressure. Arch Dermatol 1975;8:60–64
86 Kischer CW, Bunce H, Shetlar MR. Mast cell analyses in hypertrophic scars, hypertrophic scars treated with pressure and mature scars. J Invest Dermatol 1978;70:355–357
87 Sloan DF, Brown RD, Wils CH, Hilton JG. Tissue gases in human hypertrophic burn scars. Plast Reconstr Surg 1978;61:432–436
88 Shepherd JP, Dawber RF. The response of keloid scars to cryotherapy. Plast Reconstr Surg 1982;70:677–682
89 Craig RDP, Schofield JD, Jackson DS. Collagen biosynthesis in normal and hypertrophic scars and keloids as a function of the duration of the scar. Br J Surg 1975;62:741
90 Doornbos JF, Stoffel TJ, Hass AC, et al. The role of kilovoltage irradiation in the treatment of keloids. Int J Radiat Oncol Biol Phys 1990;18:833–839
91 Inalsingh CHA. An experience in treating five hundred and one patients with keloids. Johns Hopkins Med J 1974;134:284–290
92 Enhamre A, Hammar H. Treatment of keloids with excision and post-operative X-ray irradiation. Dermatologica 1983;167:90–93
93 Craig RDP, Pearson D. Early post-operative irradiation in the treatment of keloid scars. Br J Plast Surg 1965;18: 369–376
94 Hintz BL. Radiotherapy for keloid treatment. J Natl Med Assoc 1973;65:71–75
95 Ship AG, Weiss PR, Mincer FR, Wolkstein W. Sternal keloids: successful treatment employing surgery and adjunctive radiation. Ann Plast Surg 1993;31:481–487
96 Klumpar DI, Murray JC, Anscher M. Keloids treated with excision followed by radiation therapy. J Am Acad Dermatol 1994;31:225–231
97 Bilbey JH, Muller NL, Miller RR. Localized fibrous mesothelioma of pleura following external ionizing radiation therapy. Chest 1988;94:1291–1292
98 Bailin P. Use of the CO_2 laser for non-PWS cutaneous lesions. In: Arndt KA, Noe JM, Rosen S, eds. Cutaneous laser therapy: principles and methods. New York: John Wiley, 1983:187–200
99 Olbricht SM, Arndt KA. Lasers in cutaneous surgery. In: Fuller T, ed. Surgical lasers: a clinical guide. New York: MacMillan, 1987:113–146
100 Anderson RR, Parrish JA. Selective photothermolysis: precise microsurgery by selective absorption of pulsed radiation. Science 1983;22:524–527
101 Ginsbach G, Kohnel W. The treatment of hypertrophic scars and keloids by argon-laser: clinical data and morphologic findings. Plast Surg Forum 1978;1:61–67
102 Castro DJ, Abergel RP, Johnston KJ, et al. Wound healing: biological effects of Nd:YAG laser on collagen metabolism in pig skin in comparison to thermal burn. Ann Plast Surg 1983;11:131–140
103 Goldman MP, Fitzpatrick RE. Laser treatment of scars. Dermatol Surg 1995;21:685–687
104 Fulton JE. Modern dermabrasion techniques: a personal appraisal. J Dermatol Surg Oncol 1987;13:780–789
105 Ahn ST, Monafo WM, Mustoe TA. Topical silicone gel for the prevention and treatment of hypertrophic scars. Arch Surg 1991;126:499–504
106 Goldman MP, Fitzpatrick RE. Treatment of cutaneous vascular lesions. In: Goldman MP, Fitzpatrick RE, eds. Cutaneous laser surgery: the art and science of selective photothermolysis. St. Louis: Mosby, 1994:19–105

107 Nelson BR, Majmudar G, Griffiths CEM, et al. Clinical improvement following dermabrasion of photoaged skin correlates with synthesis of collagen. Arch Dermatol 1994;130:1136–1142

108 Frank W. Therapeutic dermabrasion, back to the future. Arch Dermatol 1994;130:1187–1189

109 Ellis DA, Michell MJ. Surgical treatment of acne scarring: non-linear scar revision. J Otolaryngol 1987;16:116–119

110 Johnson WC. Treatment of pitted scars: punch transplant technique. J Dermatol Surg Oncol 1986;12:260–265

111 Solotoff S. Treatment for pitted acne scarring—postauricular punch grafts followed by dermabrasion. J Dermatol Surg Oncol 1986;12:1079–1084

112 Yarborough JM, Coleman WP. Dermabrasion. In: Robinson JK, Arndt KA, LeBoit PE, Wintroub BU, eds. Atlas of cutaneous surgery. Philadelphia: WB Saunders, 1996: 333–335

113 Roenigk Jr HH. Dermabrasion: state of the art. J Dermatol Surg Oncol 1985;11:306–314

114 Orentreich N, Durr NP. Rehabilitation of acne scarring. Dermatol Clin 1983;1:405–413

115 Orentreich DS, Orentreich N. Subcutaneous incisionless (subcision) surgery for the correction of depressed scars and wrinkles. Dermatol Surg 1995;21:543–549

116 Stegman SJ, Tromovitch TA. Implantation of collagen for depressed scars. J Dermatol Surg Oncol 1980;6: 450–453

117 Klein AW. Implantation techniques for injectable collagen. J Am Acad Dermatol 1983;9:224–228

118 Varnavides CK, Forster RA, Cunliffe WJ. The role of bovine collagen in the treatment of acne scars. Br J Dermatol 1987;116:199–206

119 Selmanowitz VJ, Orentreich N. Medical grade fluid silicone: a monographic review. J Dermatol Surg Oncol 1977;3:579–611

120 Pinski KS, Roenigk HH. Autologous fat transplantation. J Dermatol Surg Oncol 1992;18:179–184

121 Spangler AS. New treatment of pitted scars. Arch Dermatol 1957;76:708–711

122 Spangler AS. Treatment of depressed scars with fibrin foam—17 year experience. J Dermatol Surg Oncol 1975; 1:65–69

123 Gold MH. The Fibrel mechanism of action study. J Dermatol Surg Oncol 1994;20:586–590

124 West TB, Alster TS. Autologous human collagen and fibroblasts for soft tissue augmentation. Dermatol Surg 1997 (submitted for publication)

125 Teimourian B. Blindness following fat injections. Plast Reconstr Surg 1988;80:361 [letter]

126 Anderson RR. Laser-tissue interactions. In: Goldman MP, Fitzpatrick RE, eds. Cutaneous laser surgery: the art and science of selective photothermolysis. St. Louis: Mosby, 1994:1–18

127 Hobbs ER, Bailin PL, Wheeland RG, Ratz JL. Superpulsed lasers: minimizing thermal damage with short duration, high irradiance pulses. J Dermatol Surg Oncol 1987;13: 955–964

128 Walsh JT, Flotte TH, Anderson RR, et al. Pulsed CO_2 laser tissue ablation: effect of tissue type and pulse duration on thermal damage. Lasers Surg Med 1988;8:108–118

129 Fitzpatrick RE, Tope WD, Goldman MP, Satur NM. Pulsed carbon dioxide laser, trichloroacetic acid, Baker-Gordon phenol, and dermabrasion: a comparative clinical and histologic study of cutaneous resurfacing in a porcine model. Arch Dermatol 1996;132:469–471

130 Alster TS, Kauvar ANB, Geronemus RG. Histology of high-energy pulsed CO_2 laser resurfacing. Semin Cutan Med Surg 1996;15:189–193

131 Smith KS, et al. Increased smooth muscle actin, factor XIIIa, and vimentin positive cells in the papillary dermis of CO_2 laser debrided porcine skin. Dermatol Surg 1997 (in press)

132 Garrett AB, Dufresne RG, Ratz JL, Berlin AJ. Carbon dioxide laser treatment of pitted acne scarring. J Dermatol Surg Oncol 1990;16:737–740

133 Ho CH, Nguyen Q, Lowe NJ, et al. Laser resurfacing in pigmented skin. J Dermatol Surg Oncol 1995;21:1035–1037

134 Apfelberg DB. A critical appraisal of high-energy pulsed carbon dioxide laser facial resurfacing for acne scars. Ann Plast Surg 1997;38:95–100

135 Alster TS, West TB. Resurfacing of atrophic facial scars with a high-energy, pulsed carbon-dioxide laser. Dermatol Surg 1996;22:151–155

136 Nanni CA, Alser TS. Cutaneous carbon dioxide laser resurfacing: long term follow-up of 300 patients. Lasers Surg Med Suppl 1997;9:39 [abstract]

137 Clark RAF. Biology of dermal wound repair. Dermatol Clin 1993;11:647–666

138 Kirsner RS, Eaglstein WH. The wound healing process. Dermatol Clin 1993;11:629–640

139 Rubenstein R, Roenigk HH, Stegman SJ, Hanke CW. Atypical keloids after dermabrasion of patients taking isotretinoin. J Am Acad Dermatol 1986;15:280–285

140 Weinstein C, Alster TS. Skin resurfacing with high energy, pulsed carbon dioxide lasers. In: Alster TS, Apfelberg DB, eds. Cosmetic laser surgery. New York: John Wiley, 1996:9–27

141 Zachariae H. Delayed wound healing and keloid formation following argon laser treatment or dermabrasion during isotretinoin treatment. Br J Dermatol 1988;118: 703–706